IMMUNOASSAYS

Use antibodies for analytical purposes

Most used type of assay that use a label to have an analytical signal

- Radio immunoassay (a radioactive label, i.e I₁₂₅ for thiroyd hormones)
- Fluorimetric assays (a fluorescent label, i.e fluorescein)
- Chemiluminometric assays (a chemiluminescent label is used)
- Enzyme immunoassays (or ELISA) (an enzyme is the label and the product of the reaction can be detected colorimetrically, fluorimetrically or by chemiluminescence)

Enzyme Linked Immuno-Sorbent Assay (ELISA)



Antibodies

•Proteins secreted by B-lymphocytes (type of white blood cell), in vertebrates.

 Recognise and bind to molecules
(antigens) on foreign particles, marking them for destruction by Tlymphocytes.

• Each antigen may generate several antibodies for different sites (**epitopes**) on antigen.





IgG molecule





Tumor necrosis factor alpha (TNF- α), also known as cachectin and TNFSF1A, is the prototypic ligand of the TNF superfamily (1). It is a pleiotropic molecule that plays a central role in inflammation, immune system development, apoptosis, and lipid metabolism (2-5). TNF- α is also involved in a number of pathological conditions including asthma, Crohn's disease, rheumatoid arthritis, neuropathic pain, obesity, type 2 diabetes, septic shock, autoimmunity, and cancer (5-11).

Human TNF-α is synthesized as a 26 kDa type II transmembrane protein that consists of a 35 amino acid (aa) cytoplasmic domain, a 21 aa transmembrane segment, and a 177 aa extracellular domain (ECD) (12, 13). Within the ECD, human TNF-α shares 97% aa sequence

This assay employs the quantitative sandwich enzyme immunoassay technique. A monoclonal antibody specific for TNF- α has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any TNF- α present is bound by the immobilized antibody. After washing away any unbound substances, an enzyme-linked polyclonal antibody specific for TNF- α is added to the wells. Following a wash to remove any unbound antibody-enzyme reagent, a substrate solution is added to the wells and color develops in proportion to the amount of TNF- α bound in the initial step. The color development is stopped and the intensity of the color is measured.

Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the TNF-a concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.



CALIBRATOR DILUENT RD6-35



Clinica Chimica Acta 510 (2020) 73-78



Comparison of test performance of commercial anti-SARS-CoV-2 immunoassays in serum and plasma samples



Verena Haselmann^{*}, Maximilian Kittel, Catharina Gerhards, Margot Thiaucourt, Romy Eichner, Victor Costina, Michael Neumaier

Department of Clinical Chemistry, University Medicine Mannheim, Medical Faculty Mannheim of the Univ

At the end of 2020, over 100 SARS-CoV-2 antibody assays have been CE-marked under EU Directive 98/79/EC . The available test systems can be discriminated into rapid diagnostic tests (RDT), either antigen- or antibody based, enzyme-linked immunosorbent assays (ELISA) and chemiluminescent immunoassays (CLIA).

Journal of Clinical Virology 139 (2021) 104821





Head-to-head validation of six immunoassays for SARS-CoV-2 in hospitalized patients

Rens Zonneveld ^{a, *, 1}, Suzanne Jurriaans ^{a, 1}, Tom van Gool ^a, Jorrit J. Hofstra ^a, Thecla A. M. Hekker ^a, Pien Defoer ^a, Patricia E. Broekhuizen-van Haaften ^a, Ellen M. Wentink-Bonnema ^a, Lynn Boonkamp ^b, Charlotte E. Teunissen ^b, Annemieke C. Heijboer ^c, Frans Martens ^c, Godelieve de Bree ^d, Michele van Vugt ^d, Robin van Houdt ^a, Amsterdam UMC COVID-19 Biobank



Magnetic Microbeads are very often used for the separation step in immunoasssays



A chemiluminescence immunoassay principle in

Chemical reaction of Assay scheme the light emission

Main Reaction Components



Competitive immunoassays



(b)

$$y = \frac{a - d}{(1 + (x/c)^{b})} + d$$

- a = (theoretical) response at low concentration/dilution
- b = absolute value of the slope at the inflection point
- c = value of x at inflection point
- d = (theoretical) response at high concentration/dilution
- x = concentration or dilution

y = response (OD)

Typical calibration curve for immunoassay



Figure 1. Typical 4-parameter logistic graph for a competitiveformat immunoassay.

Enzymes of ELISA

Enzyme*	Source	Specific Enzyme activity (units∆/mg)
Alkaline phosphatase	Calf intestine	400
Beta Galactosidase	E.coli	400
Glucose oxidase	Aspergillus Niger	200
Glucose -6-phosphate dehydrogenase	Leucon Stoc mesenteroides	250
Peroxidase	Horseradish	900

ENZYME SUBSTRATE

- Initially the substrate should be colorless
- After degradation by the enzyme it should be strongly colored or fluorescent.

ENZYME	SUBSTRATE	CHROMOGEN	STOPPING
Alkaline Phosphatase	p-NPP	p-NPP+ diethandamine+Mg Cl2	I M NaOH
Horse radish Peroxidase	H2O2	Tetramethylbenzidi ne + Phosphate – Citrate buffer	I M H2SO4
Horse radish Peroxidase	H2O2	0 – Phenylenediamine + HCl	I M HCI
			14



Horseradish peroxidase and alkaline phosphatase retain high activity when conjugated with antibodies or antigens and have different types of substrates useful for different assays





À la carte ELISA Systems

Almond	Beta- Lactoglobulin	Buckwheat	Casein	Crustacean
			55538 S S S S S S	
Egg	Gluten	Hazelnut	Lupin	Mustard
Peanut	Sesame	Sov		
	Almond Egg Peanut	Almond Beta- Lactoglobulin Egg Gluten Peanut Sesame	AlmondBeta-LactoglobulinBuckwheatEggGlutenHazelnutEggSesameSoy	AlmondBeta- LactoglobulinBuckwheatCaseinBuckwheatCaseinImage: Second State Stat

Polyclonal antibodies production



Monoclonal antibodies



Recombinant antibodies



Fig. 1. Illustration of scFv library generation.



ELISA

• Enzyme-linked immunosorbent assay (ELISA) is a test that uses antibodies and color change to identify a substance.





Electrochemical immunosensors (labeled)









$f(x) = \{(a - d)/[1 + (x/c)b]\} + d$

	Col	Linear regression			
	a (A or nA)	b (nA.ng.ml ⁻¹)	c (ng.ml ⁻¹)	d (A or nA)	
ic spettr	1.220 ± 0.053	1.40 ± 0.50	0.80 ± 0.22	0.129 ± 0.077	$f(x) = 49.3 (\pm 0.8) - 57.5 (\pm 0.1) x$ [r = 0.991]
ic amp.	6019 ± 118	0.90 ± 0.22	0.93 ± 0.10	176 ± 30	$f(x) = 52.5 (\pm 0.4) - 43.7 (\pm 0.5) x$ $[r = 0.994]$
dc spettr	1.392 ± 0.061	2.17 ± 0.15	0.80 ± 0.14	0.132 ± 0.071	$f(x) = 47.4 (\pm 0.7) - 86.0 (\pm 0.3) x$ $[r = 0.993]$
dc amp.	707 ± 56	1.10 ± 0.10	0.35 ± 0.04	16±13	$f(x) = 34.9 (\pm 0.6) - 52.2 (\pm 0.9) x$ $[r = 0.992]$

Immunoassay	Working Range (ng/ml)	L.O.D. (Blank – 3 σ) (ng/ml)
ic spettr	0.20 – 2.5	0.150
ic amp.	0.10 – 7.5	0.120
dc spettr	0.10 - 10	0.080
dc amp.	0.05 - 2.5	0.060

Immunosensor procedure:

✓ Pre-coating: 6 μ l of rabbit anti IgG (4° C overnight)

✓Blocking: 6 μ l of 1 % PVA (polyvinyl alcohol) (30 min)

 \checkmark Coating: 6 μl of anti-OTA Ab (1 h)

 \checkmark Competition: 6 µl of OTA-AP + standard/sample (30 min)

✓Detection: 100 of 5 mg/ml 1-Naphtylphosfate (2 min) + DPV

washings: 150 μl phosphate buffer pH 7.4

effect of extraction solvent

Activity of an electrode modified with IgG-ALP after 30 min incubation with 1:9 -9:1 solutions (1:1 in DPBS) acetonitrile:water \rightarrow 95-108%

Sensitivity of the calibration curve ~ 50%





25 g in 100 mL di ACN:H20

Final dilution 1:8

Maximum Residue Limit = 3 ng/g

 I_{50} = 1.6 ng/g

Paramet	ers	0.1 PBS Buffer •	ACN:H ₂ O (6:4) ▲	Wheat Extract (blank) ■
а	(nA)	707 (± 56)	260 (± 12)	408 (± 72)
b	(nA.ng.ml ⁻¹)	1.1 (± 0.1)	0.62 (± 0.03)	0.8 (± 0.1)
c (I ₅₀)	(ng.ml ⁻¹)	0.35 (± 0.04)	0.32 (± 0.02)	0.20 (± 0.03)
d	(nA)	16 (± 13)	24 (± 8)	13 (± 15)
w.r.	(ng.ml ⁻¹)	0.05 - 2.5	0.02 - 5.0	0.05 - 2.5
L.O.D.	(ng.ml ⁻¹)	0.06	0.015	0.05
Lin.	Reg.	$30.9 (\pm 0.6) - 52.2 (\pm 0.9) x$	42.3 (± 0.3) – 25.4 (± 0.6) x	23.5 $(\pm 0.1) - 41.1 \ (\pm 0.5) \ x$

Lateral flow immunoassays













SPR Biosensors label-free!!







- dextran hydrogel open structure (good accessibility) no denaturation
- enhancement of the capacity of the interaction layer
- stagnant layer / mass transport flow needed (µl/min)
- negative charge
- regenerable (up to 100 x)

SPR principles

Surface plasmon resonance (SPR) arises when light is reflected under certain conditions from a conducting film at the interface between two media of different refractive index. the media are the <u>sample</u> and the <u>glass</u> of the sensor chip, and the conducting film is a thin layer of gold on the chip surface. SPR causes a reduction in the intensity of reflected light at a specific angle of reflection. This angle varies with the refractive index close to the surface on the side opposite from the reflected light.

When molecules in the sample bind to the sensor surface, the concentration and therefore the refractive index at the surface changes and an SPR response is detected. Plotting the response against time during the course of an interaction provides a quantitative measure of the progress of the interaction. This plot is called a sensorgram.

What Biacore actually measures is the angle of minimum reflected light intensity. The light is not absorbed by the sample: instead the light energy is dissipated through SPR in the gold film. Thus the light used to detect interaction processes never enters the sample.

SPR response values are expressed in resonance units (RU). One RU represents a change of 0.0001° in the angle of the intensity minimum. For most proteins, this is roughly equivalent to a change in concentration of about 1 pg/mm2 on the sensor surface. The exact conversion factor between RU and surface concentration depends on properties of the sensor surface and the nature of the molecule responsible for the concentration change.





Immobilisation of organic molecules on gold







BIO(immuno)FETs label-free!

An **ion-sensitive field-effect transistor** (**ISFET**) is a <u>field-effect transistor</u> used for measuring ion concentrations in solution; when the ion concentration (such as <u>H</u>⁺, see <u>pH</u> scale) changes, the current through the <u>transistor</u> will change accordingly. Here, the solution is used as the gate electrode. A voltage between substrate and <u>oxide</u> surfaces arises due to an <u>ion</u> sheath. It is a special type of <u>MOSFET</u> (metaloxide-semiconductor field-effect transistor),^[1] and shares the same basic structure, but with the <u>metal gate</u> replaced by an ion-sensitive <u>membrane</u>, <u>electrolyte</u> solution and <u>reference electrode</u>.^[2] Invented in 1970, the ISFET was the first <u>biosensor</u> <u>FET</u> (BioFET) source wikipedia



Bio-FETs couple a <u>transistor</u> device with a bio-sensitive layer that can specifically detect bio-molecules such as nucleic acids and proteins. A Bio-FET system consists of a semiconducting <u>field-effect transistor</u> that acts as a <u>transducer</u> separated by an insulator layer (e.g. <u>SiO₂</u>) from the biological recognition element (e.g. receptors or probe molecules) which are selective to the target molecule called analyte.^[8] Once the analyte binds to the recognition element, the charge distribution at the surface changes with a corresponding change in the electrostatic surface potential of the semiconductor. This change in the surface potential of the semiconductor acts like a gate voltage would in a traditional <u>MOSFET</u>, i.e. changing the amount of current that can flow between the source and drain electrodes.^[9] This change in current (or <u>conductance</u>) can be measured, thus the binding of the analyte can be detected. The precise relationship between the current and analyte concentration depends upon the <u>region of transistor</u> <u>operation</u> (source Wikipedia)





<u>Piezoelectric Biosensors label-free!</u>













The mass-loading frequency effects of the transducer are based upon Sauerbrey's equation



The standard QCM measures the mass of a material deposited on a quartz crystal surface as a linear function of a change in the oscillating crystal resonant frequency International Journal of ELECTROCHEMICAL SCIENCE www.electrochemsci.org

Short Communication

Piezoelectric Immunosensor for the Determination of C-Reactive Protein

Miroslav Pohanka

Faculty of Military Health Sciences, University of Defence, Trebesska 1575, CZ-500 01 Hradec Kralove, Czech Republic E-mail: <u>miroslav.pohanka@gmail.com</u>

C-Reactive protein is a marker of inflammation



Figure 1. Principle of CRP assay by the proposed biosensor.



Figure 2. Calibration of CRP using piezoelectric biosensor in combination with modified nanoparticles as a label.



Figure 3. Bar graph describing effect of potential interferents albumin 100 mg/ml (bar 3), IgM 50 mg/ml (bar 4) and TNFα 100 ng/ml (bar 5) on biosensor serving for CRP assay. CRP in concentration 12.5 mg/l (bar 1) and pure PBS (bar 2) are given for comparison.

