

# 25 DNA Metabolism

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Learning



# Principle 1 (1 of 6)

**Along with catalysis, biological information is one of the two key prerequisites for life.** The faithful maintenance and transmission of genetic information from one generation to another ensures continuity within each species.

## Principle 2 (1 of 7)

**Information is expensive.** The chemistry of joining one nucleotide to the next in DNA replication is elegant and simple, almost deceptively so. However, the enzymatic and thermodynamic commitment to linking one nucleotide to another in DNA far exceeds what would normally be required to successfully form a phosphodiester bond. It is not enough to synthesize a phosphodiester bond; that bond must accurately link two *particular* nucleotides.

## Principle 3 (1 of 7)

**The fidelity of genome maintenance and transmission is not perfect.** DNA damage happens, often by spontaneous processes. DNA replication and repair deal with the vast majority of DNA lesions, providing a high degree of genetic fidelity and stability. The few DNA damage events that slip through uncorrected provide fuel for evolution.

## Principle 4 (1 of 5)

**Although considered separately, the processes of replication, repair, and recombination of DNA are not distinct.** These processes are highly integrated in cells, and they are required for proper genome maintenance.

# Bacterial Gene Naming

- typically named using three italicized lowercase letters reflecting a function
  - examples: *dna*, *uvr*, and *rec*
- capital letters added to abbreviation reflect order of discovery, not enzymatic order
  - examples: *dnaA*, *dnaB*, and *dnaQ*

# Bacterial Protein Naming

- often named after their genes using nonitalicized, roman type with the first letter capitalized
  - examples: DnaA (encoded by *dnaA*) and RecA (encoded by *recA*)

# Eukaryotic Gene Naming

- no single convention exists for all eukaryotic systems
- in *Saccharomyces cerevisiae*, gene names are three italicized uppercase letters followed by an italicized number
  - example: *COX1*

# Eukaryotic Protein Naming

- complex and variable
- in yeast, some proteins have long common names
  - example: cytochrome oxidase
- other yeast proteins have the same name as the gene, with one uppercase and two lowercase letters in roman type, followed by a number and the letter “p”
  - example: Rad51p (encoded by *RAD51*)

# *E. Coli* Has at Least Five DNA Polymerases

- DNA polymerase I is abundant, but insufficient for replication of the *E. coli* chromosome
  - rate (600 nucleotides/min) is slower than observed for replication fork movement
  - low processivity
- the primary function of DNA polymerase I is cleanup during replication, recombination, and repair

# DNA Polymerase II, III, IV, and V

- **DNA polymerase II** = involved in DNA repair
- **DNA polymerase III** = the principal replication enzyme in *E. coli*
- DNA polymerases IV and V = involved in an unusual form of DNA repair

# Comparison of the Five DNA Polymerases of *E. coli*

**Table 25-1 Comparison of the Five DNA Polymerases of *E. coli***

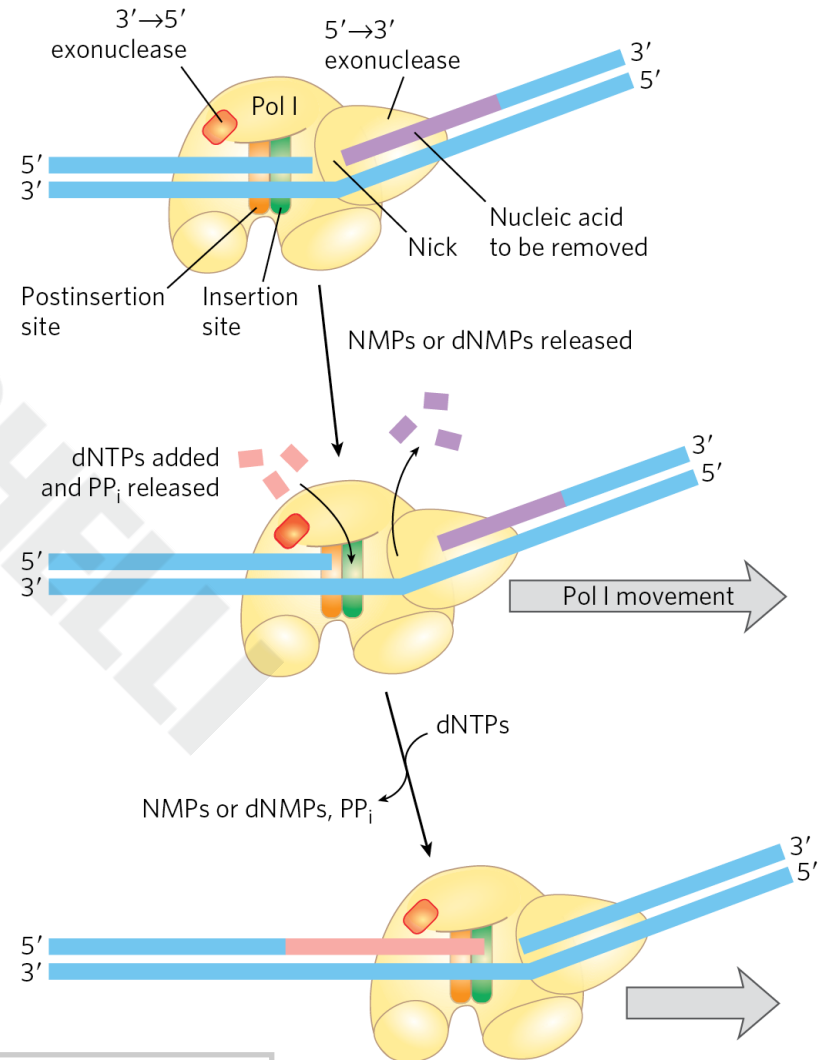
	DNA polymerase I	DNA polymerase II	DNA polymerase III	DNA polymerase IV	DNA polymerase V
Structural gene <sup>b</sup>	<i>polA</i>	<i>polB</i>	<i>polC (dnaE)</i>	<i>dinB</i>	<i>umuC</i>
Subunits (number of different types)	1	7	9	1	3
$M_r$	103,000	88,000 <sup>c</sup>	1,065,400	39,100	110,000
3'→5' exonuclease (proofreading)	Yes	Yes	Yes	No	No
5'→3' exonuclease	Yes	No	No	No	No
Polymerization rate (nucleotides/s)	10–20	40	250–1,000	2–3	1
Processivity (nucleotides added before polymerase dissociates)	3–200	1,500	≥500,000	1	6–8

# DNA Polymerase I Has 5'→3' Exonuclease Activity

- distinct from the 3'→5' proofreading exonuclease
- the 5'→3' domain is in front of the enzyme and performs nick translation
- mild protease treatment separates this domain from the remainder of the enzyme (the **large fragment** or **Klenow fragment**)

# Nick Translation

- nick translation = a break or nick in the DNA is moved along with the enzyme
- important in:
  - DNA repair
  - the removal of RNA primers during replication



# Subunits of DNA Polymerase III of *E. coli*

**TABLE 25-2** Subunits of DNA Polymerase III of *E. coli*

Subunit	Number of subunits per holoenzyme	$M_r$ of subunit	Gene	Function of subunit	
$\alpha$	3	129,900	<i>polC (dnaE)</i>	Polymerization activity	} Core polymerase
$\epsilon$	3	27,500	<i>dnaQ (mutD)</i>	3' $\rightarrow$ 5' proofreading exonuclease	
$\theta$	3	8,600	<i>holE</i>	Stabilization of $\epsilon$ subunit	
$\tau$	3	71,100	<i>dnaX</i>	Stable template binding; core enzyme dimerization	} Clamp-loading ( $\gamma$ ) complex that loads $\beta$ subunits on lagging strand at each Okazaki fragment <sup>a</sup>
$\delta$	1	38,700	<i>holA</i>	Clamp opener	
$\delta'$	1	36,900	<i>holB</i>	Clamp loader	
$\chi$	1	16,600	<i>holC</i>	Interaction with SSB	
$\psi$	1	15,200	<i>holD</i>	Interaction with $\tau$ and $\chi$	
$\beta$	6	40,600	<i>dnaN</i>	DNA clamp required for optimal processivity	

<sup>a</sup>The  $\gamma$  subunit is encoded by a portion of the gene for the  $\tau$  subunit (*dnaX*), such that the amino-terminal 66% of the  $\tau$  subunit has the same amino acid sequence as the  $\gamma$  subunit. The  $\gamma$  subunit is generated by a translational frameshifting mechanism (p. 1013) that leads to premature translational termination. The  $\gamma$  subunit shares the clamp-loading functions of  $\tau$  but lacks the protein segments that interact with the core polymerase or with DnaB helicase. Clamp-loading complexes incorporating  $\gamma$  subunits may operate independently of the DNA polymerase III holoenzyme, promoting the unloading of  $\beta$  clamps discarded on the lagging strand as the replication fork progresses. They may also promote loading of  $\beta$  clamps for some DNA repair processes that require DNA synthesis away from the replication fork.

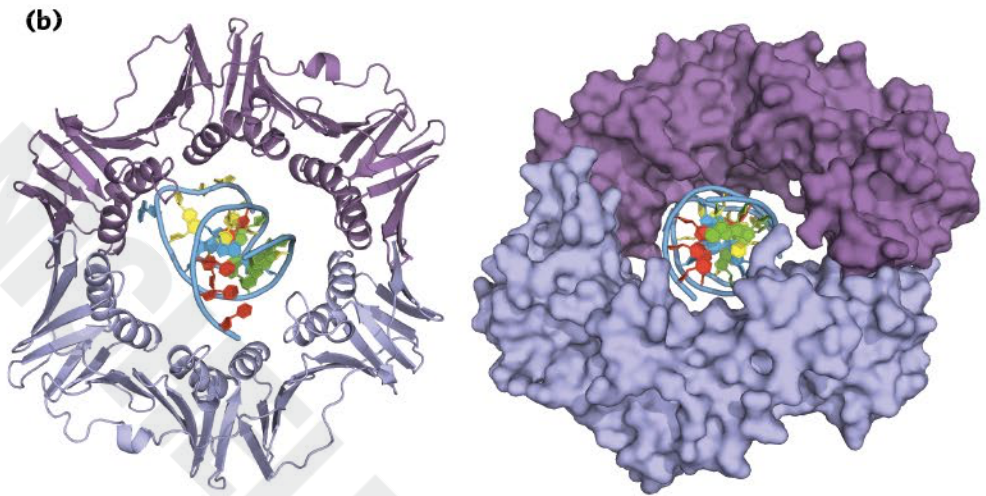
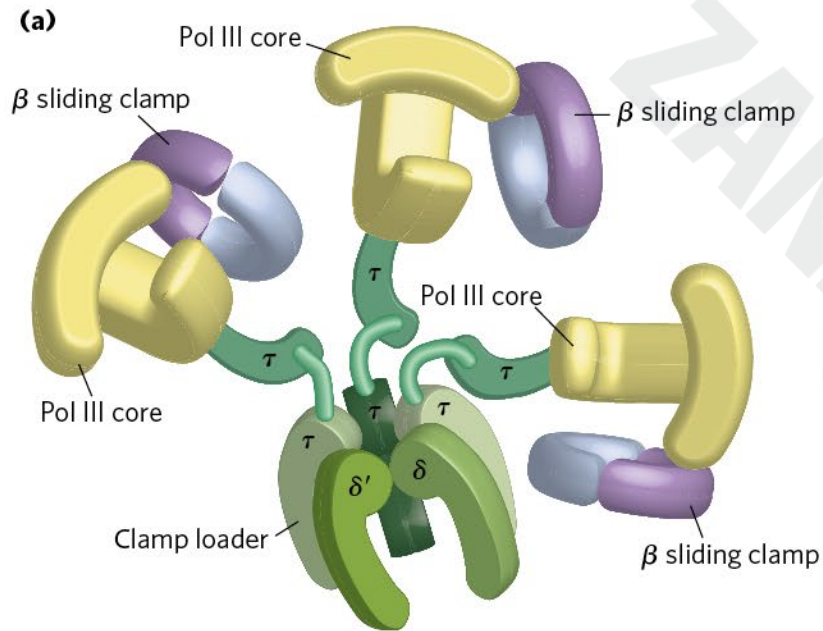
# Subunits of DNA Polymerase III\* in *E. coli*

- $\alpha$  subunit = contains polymerization activity
- $\epsilon$  subunit = contains proofreading activity
- $\theta$  subunit associates with  $\alpha$  and  $\epsilon$  subunits to form a core polymerase
  - can polymerize DNA but with limited processivity
  - up to three core polymerases are linked by a clamp-loading complex ( $\tau_3\delta\delta'$ )
- $\chi$  and  $\psi$  bind to the clamp-loading complex

# $\beta$ Subunits Converts DNA Polymerase III\* to DNA Polymerase III Holoenzyme

- DNA polymerase III\* = the entire assembly of 16 protein subunits (eight different types)
- $\beta$  subunits = provide an increase in processivity by preventing dissociation of DNA polymerase III from DNA
  - one dimer of  $\beta$  subunits associates with each active core subassembly
  - addition of  $\beta$  subunits converts DNA polymerase III\* to DNA polymerase III holoenzyme

# DNA Polymerase III



# DNA Replication Requires Many Enzymes and Protein Factors

- **DNA replicase system (replisome)** = the entire complex of enzymes and proteins required for replication in *E. coli*
  - consists of 20+ different enzymes and proteins



## Principle 2 (4 of 7)

**Information is expensive.** The chemistry of joining one nucleotide to the next in DNA replication is elegant and simple, almost deceptively so. However, the enzymatic and thermodynamic commitment to linking one nucleotide to another in DNA far exceeds what would normally be required to successfully form a phosphodiester bond. It is not enough to synthesize a phosphodiester bond; that bond must accurately link two *particular* nucleotides.

# Main Classes of Replication Enzymes

- **helicases** = enzymes that move along the DNA and separate the strands
  - requires chemical energy from ATP
- **topoisomerases** = relieve topological stress created by strand separation
- **DNA-binding proteins** = stabilize separated strands
- **primases** = synthesize short segments of RNA to serve as primers

# Main Classes of Replication Enzymes, Continued

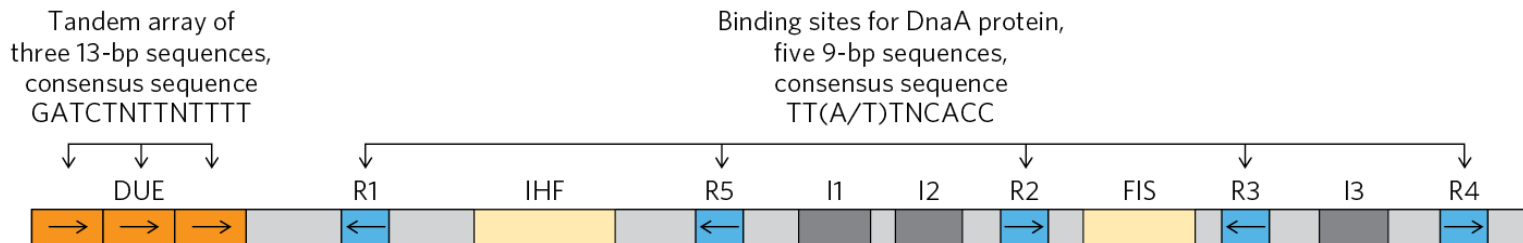
- DNA polymerase I = removes and replaces RNA primers
- RNase H1 = a specialized nuclease that degrades RNA in RNA-DNA hybrids
  - can remove RNA primers
- **DNA ligases** = seals nicks in the DNA backbone following removal and replacement of an RNA primer

# Replication of the *E. coli* Chromosome Proceeds in Stages

- DNA synthesis can be divided into three stages:
  - initiation
  - elongation
  - termination

# Initiation

- the *E. coli* replication origin (*oriC*) consists of:
  - five repeats of a 9 bp sequence (R sites) that serve as binding sites for the key initiator protein, DnaA
  - the **DNA unwinding element (DUE)** = a region rich in A=T base pairs
  - three additional DnaA-binding sites (I sites)
  - binding sites for the proteins IHF (integration host factor) and FIS (factor for inversion stimulation)



# Proteins Required to Initiation Replication at the *E. coli* Origin

**Table 25-3 Proteins Required to Initiate Replication at the *E. coli* Origin**

Protein	$M_r$	Number of subunits	Function
DnaA protein	52,000	1	Recognizes <i>oriC</i> sequence; opens duplex at specific sites in origin
DnaB protein (helicase)	300,000	6	Unwinds DNA
DnaC protein	174,000	6	Required for DnaB binding at origin
HU	19,000	2	Histonelike protein; DNA-binding protein; stimulates initiation
FIS	22,500	2	DNA-binding protein; stimulates initiation
IHF	22,000	2	DNA-binding protein; stimulates initiation
Primase (DnaG protein)	60,000	1	Synthesizes RNA primers
Single-stranded DNA-binding protein (SSB)	75,600	4	Binds single-stranded DNA
DNA gyrase (DNA topoisomerase II)	400,000	4	Relieves torsional strain generated by DNA unwinding
Dam methylase	32,000	1	Methylates (5')GATC sequences at <i>oriC</i>

# The AAA+ ATPase Protein Family

- **AAA+ ATPase** protein family (ATPases associated with diverse cellular activities) = proteins that form oligomers and hydrolyze ATP relatively slowly
  - ATP hydrolysis mediates interconversion of the protein between two states

# The DnaA Protein

- DnaA protein = a crucial component in the initiation process
  - a member of the AAA+ ATPase family
  - active when ATP is bound and inactive when ADP is bound

# DnaA Proteins Bind at R and I Sites in *oriC*

- eight ATP-bound DnaA molecules bind to the R and I sites in *oriC*
- strand separation in the A=T-rich DUE occurs due to:
  - strain due to positive supercoiling
  - binding of DnaA to the DUE region

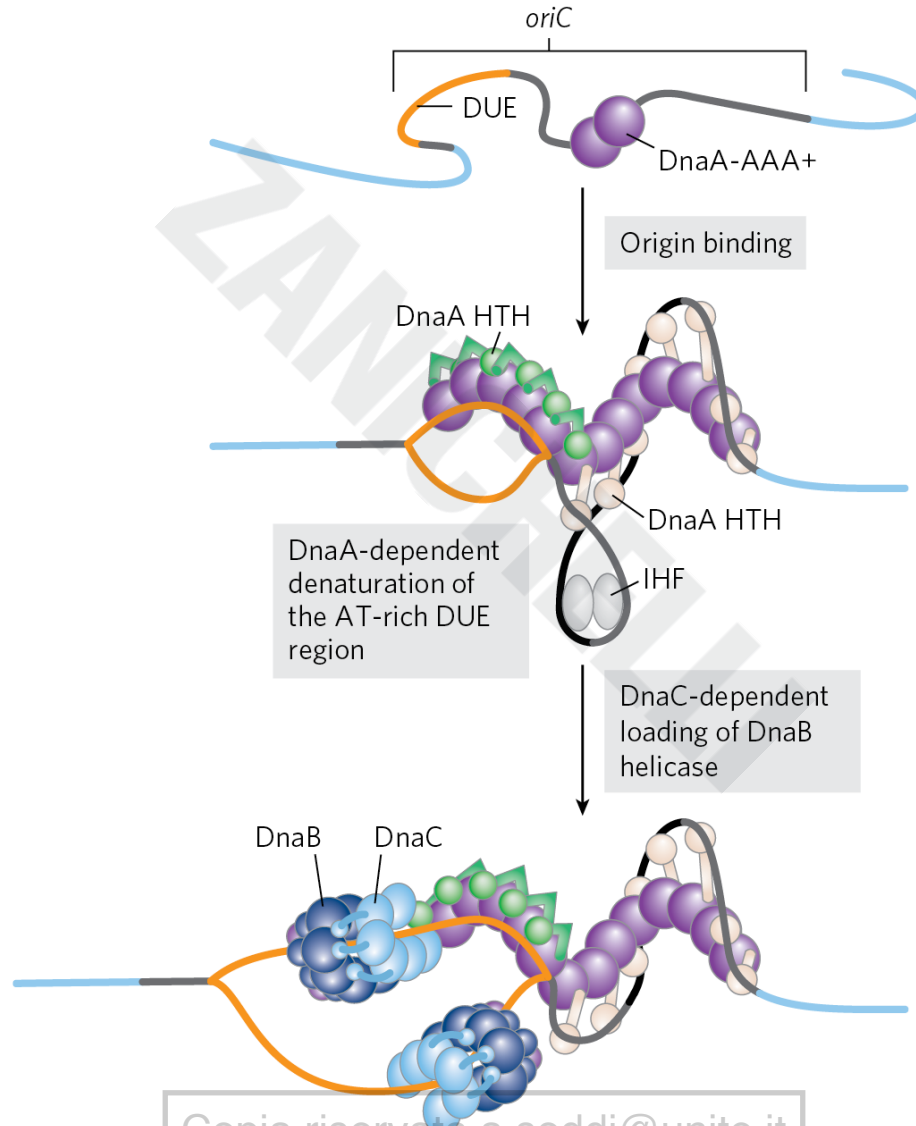
# Loading of the DnaB Helicase

- DnaC protein = an AAA+ ATPase that loads DnaB protein onto the separated DNA strands
  - two ring-shaped DnaB hexamers are loaded in the DUE, one onto each DNA strand
- DnaB protein (helicase) = migrates along ssDNA in the 5'→3' direction and unwinds DNA

# The Role of SSB and DNA Gyrase in Initiation

- single-stranded DNA-binding protein (SSB) stabilizes separated strands
- DNA gyrase relieves topological stress ahead of the replication forks

# Model for Initiation of Replication



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# Initiation Occurs Only Once in Each Cell Cycle

- Hda protein = an AAA+ ATPase that binds to the  $\beta$  subunits of DNA polymerase III and stimulates hydrolysis of the ATP bound to DnaA
  - causes disassembly of the DnaA complex
  - homologous to DnaA
- release of ADP by DnaA and rebinding of ATP occurs on a time scale of 20 to 40 minutes

# Regulation of Replication Initiation via Methylation

- Dam (*DNA adenine methylation*) methylase = methylates the  $N^6$  position of adenine within the palindromic sequence (5')GATC of *oriC* DNA
- hemimethylated *oriC* sequences are sequestered by:
  - interaction with the plasma membrane
  - binding of the protein SeqA
- after SeqA dissociation, *oriC* sequences are released from the membrane and Dam methylase fully methylates DNA to allow new DnaA to bind

# Elongation

- elongation includes:
  - leading strand synthesis
  - lagging strand synthesis

# Elongation of the Leading Strand

- the more straightforward of the two strands
- primase (DnaG) synthesizes a short (10 to 60 nucleotide) RNA primer at the replication origin
  - requires interaction with DnaB helicase
  - primase and DnaB move in opposite directions
- DNA polymerase III adds nucleotides to the 3' of the primer
  - linked to the DnaB tethered to the opposite DNA strand

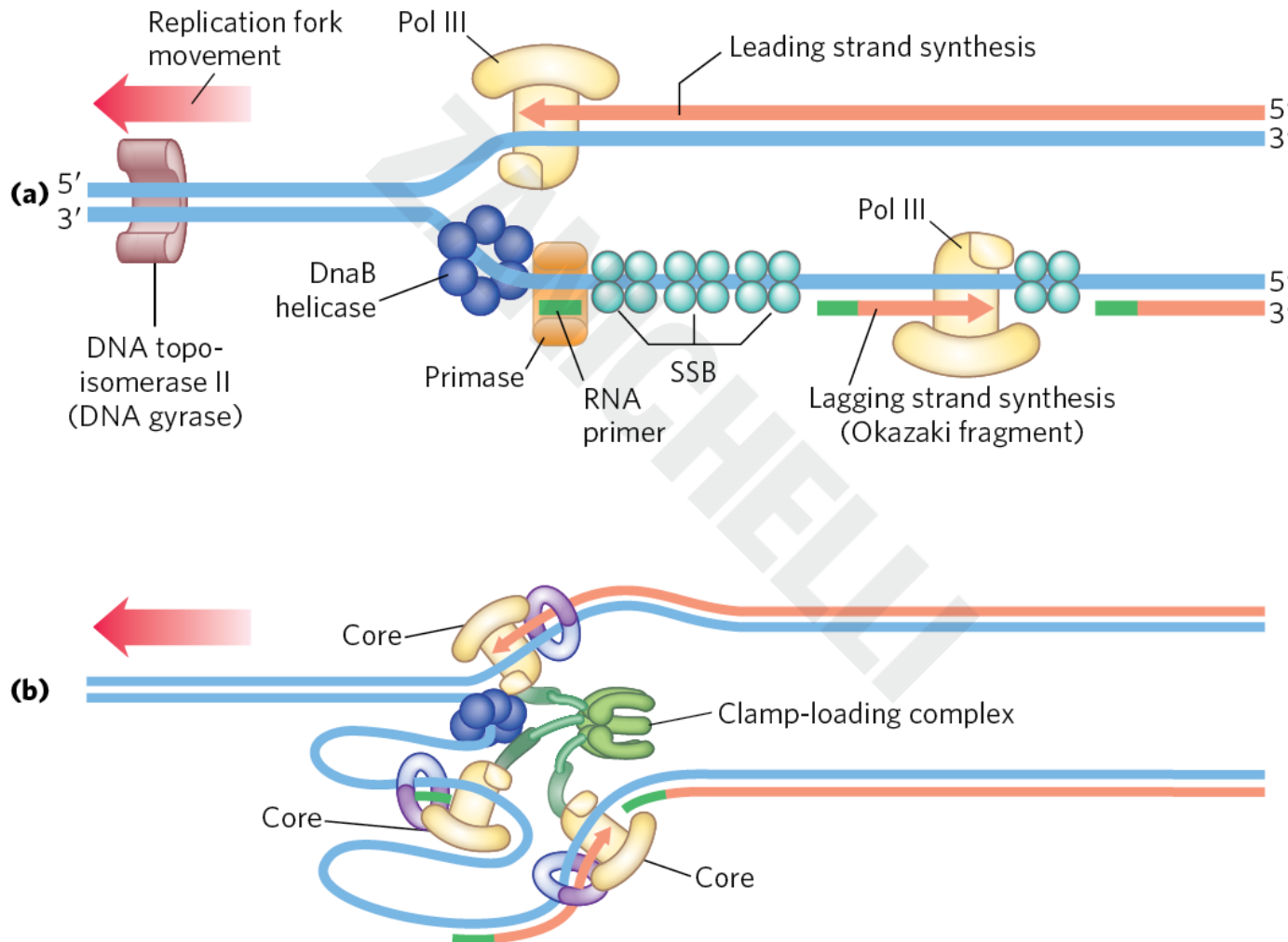
# Elongation of the Lagging Strand

- accomplished in short Okazaki fragments
- as in leading strand synthesis, primase synthesizes an RNA primer and DNA Pol III adds nucleotides to the 3' end
- DNA of the lagging strand loops around so one asymmetric DNA Pol III dimer complex can synthesize both strands

# Leading and Lagging Strand Synthesis by DNA Polymerase III

- DNA polymerase III uses one set of its core subunits to synthesize the leading strand continuously
- the other two sets of core subunits cycle from one Okazaki fragment to the next on the looped lagging strand

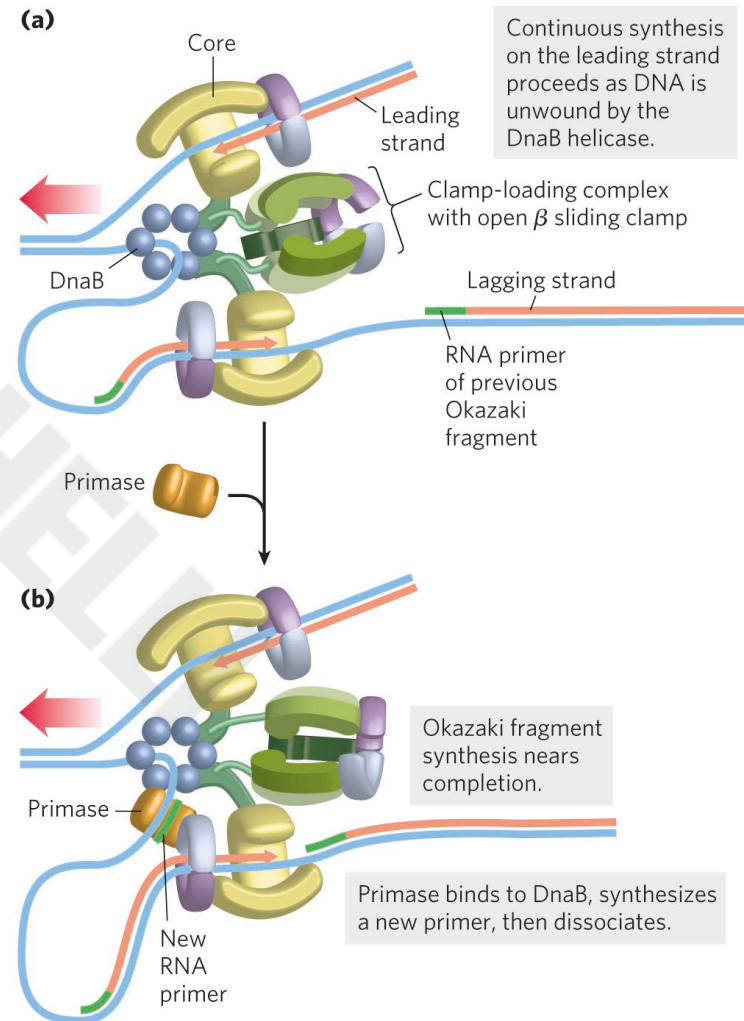
# Synthesis of Okazaki Fragments



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# Leading and Lagging Strand Synthesis, Part 1

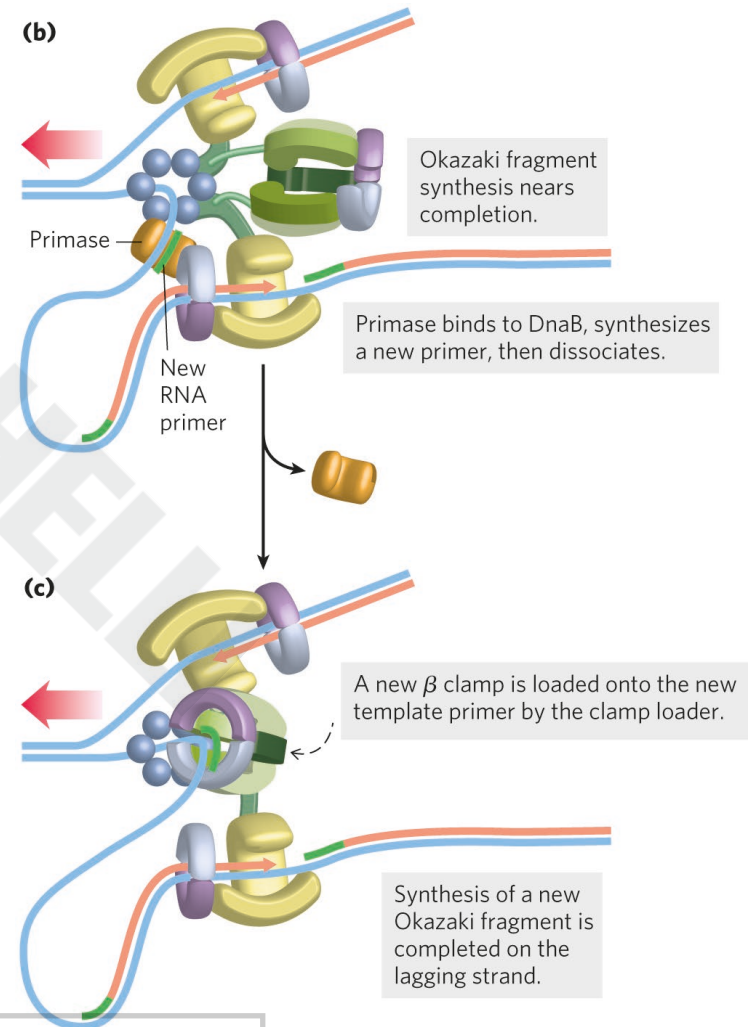
- DnaB helicase travels along the lagging strand template in the 5'→3' direction and unwinds the DNA
- DnaG primase occasionally associates with DnaB helicase and synthesizes a short RNA primer



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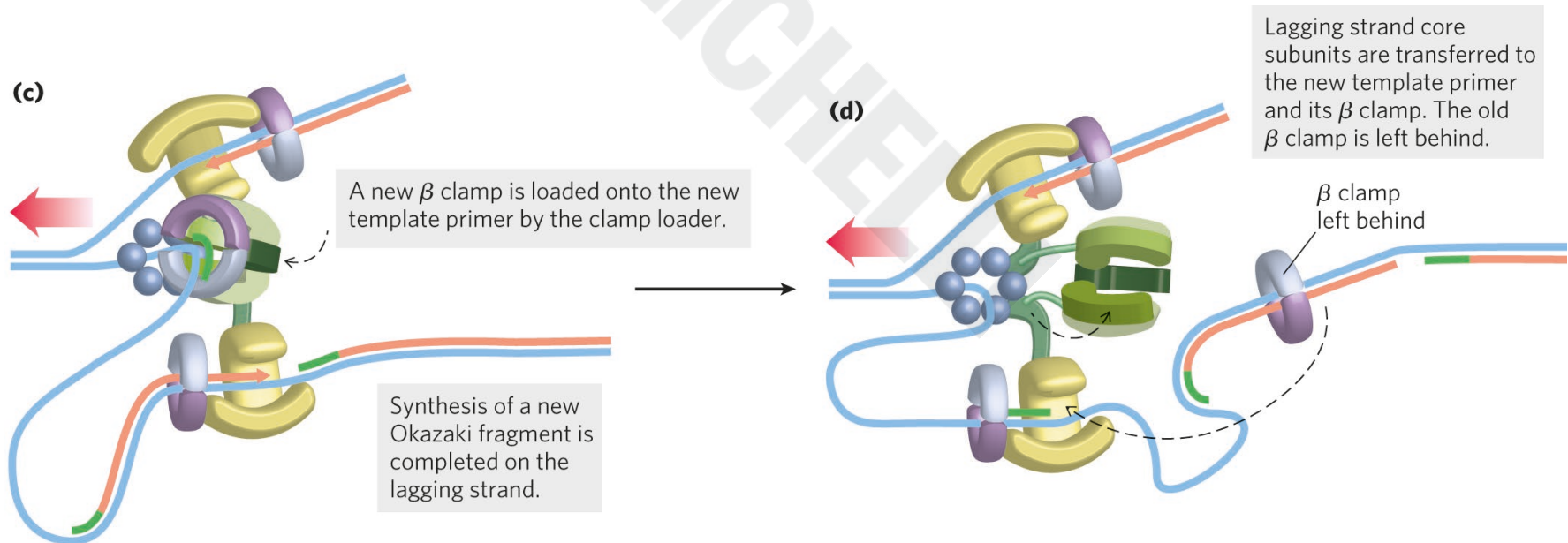
# Leading and Lagging Strand Synthesis, Part 2

- a new  $\beta$  sliding clamp is positioned at the primer by the clamp-loading complex of DNA polymerase III



# Leading and Lagging Strand Synthesis, Part 3

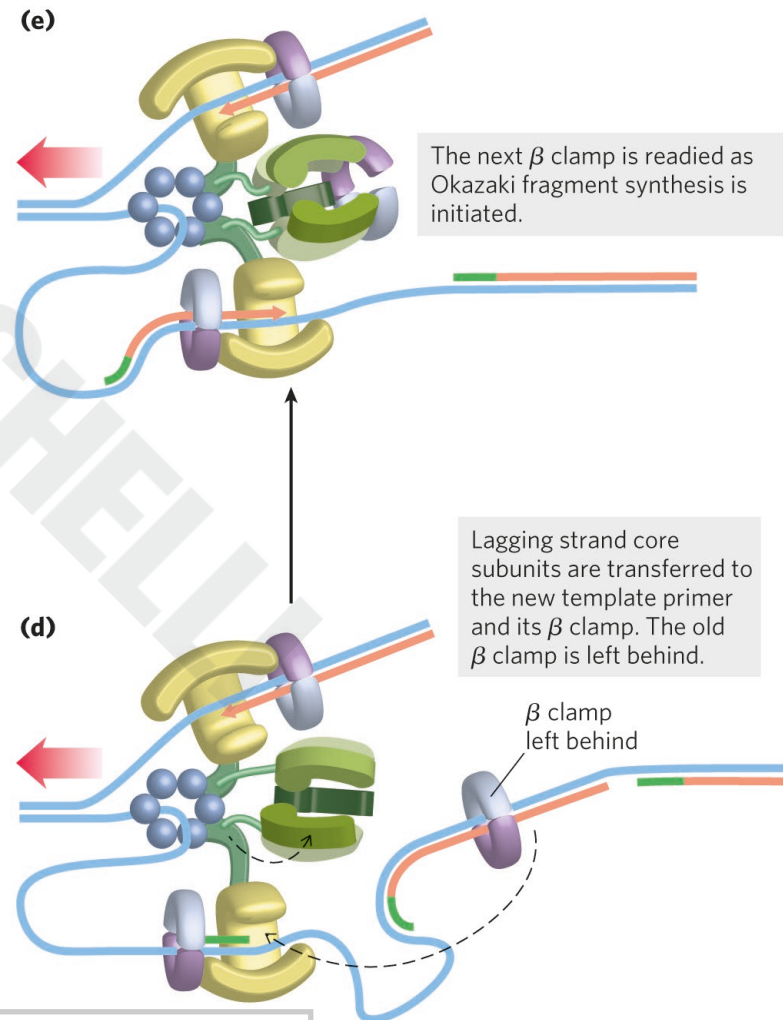
- replication halts after synthesis of an Okazaki fragment is completed and DNA polymerase III core subunits dissociate from the  $\beta$  sliding clamp



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# Leading and Lagging Strand Synthesis, Part 4

- DNA polymerase III core subunits associate with a new  $\beta$  sliding clamp
  - initiates synthesis of a new Okazaki fragment



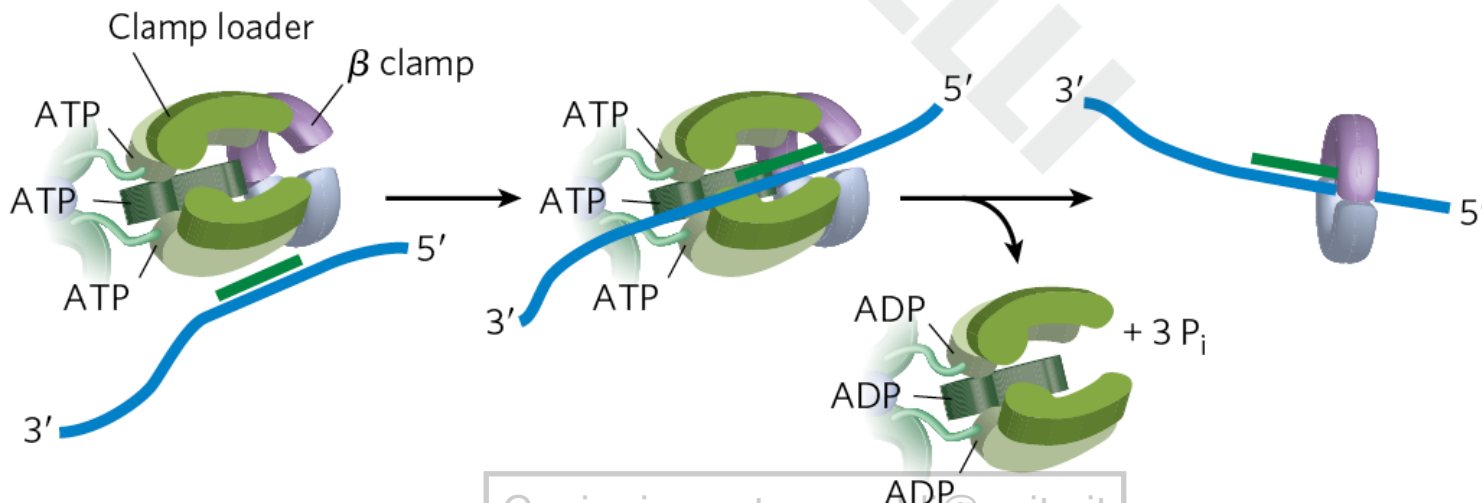
# Proteins Acting at the Replication Fork

**Table 25-4 Proteins of the *E.coli* Replisome**

Protein	$M_r$	Number of subunits	Function
SSB	75,600	4	Binding to single-stranded DNA
Helicase (DnaB protein)	300,000	6	DNA unwinding
Primase (DnaG protein)	60,000	1	RNA primer synthesis
DNA polymerase III	1,065,400	17	New strand elongation
DNA polymerase I	103,000	1	Filling of gaps; excision of primers
DNA ligase	74,000	1	Ligation
DNA gyrase (DNA topoisomerase II)	400,000	4	Supercoiling

# The DNA Polymerase III Clamp Loader

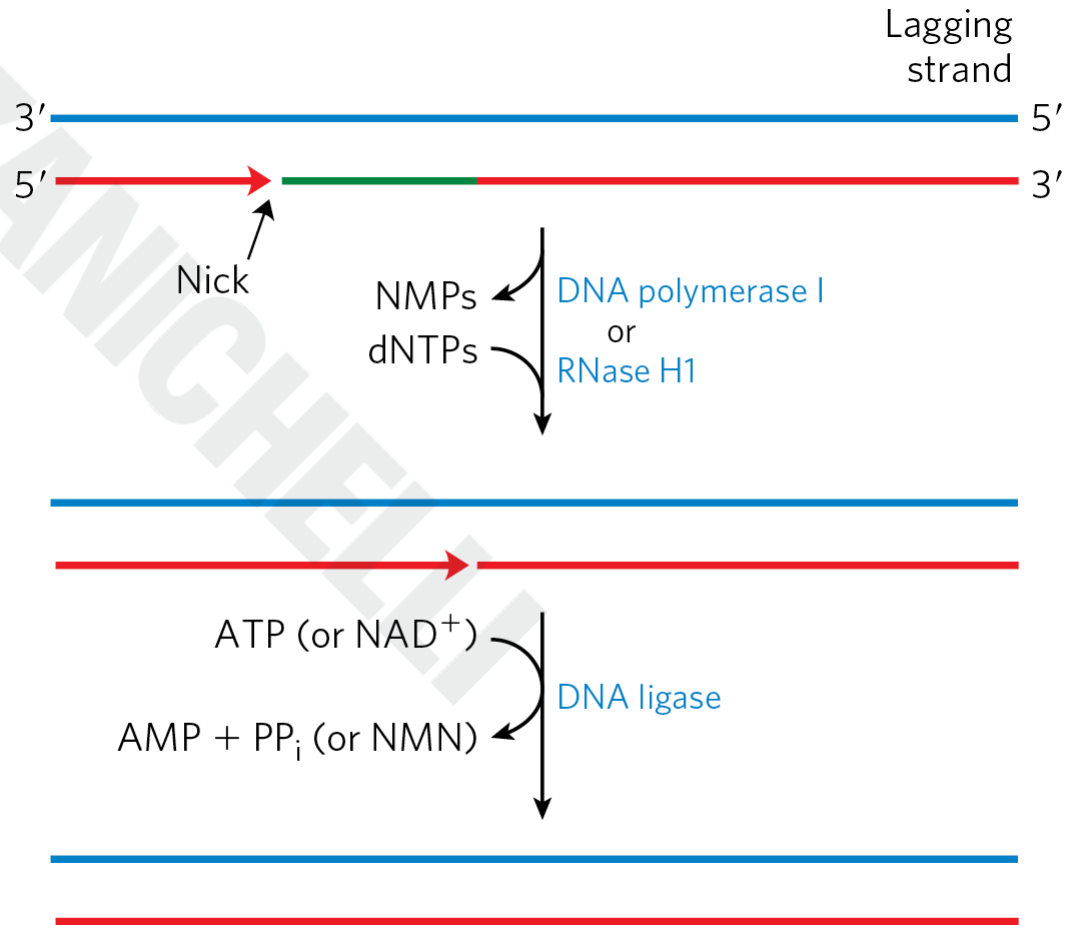
- clamp-loading complex = and AAA+ ATPase that binds ATP and the new  $\beta$  sliding clamp
  - consists of parts of the three  $\tau$  subunits along with the  $\delta$  and  $\delta'$  subunits
  - ATP binding opens the ring at one subunit interface
  - ATP hydrolysis closes the clamp around the DNA



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# The Final Steps in Lagging Strand Synthesis

- DNA polymerase I or RNase H1 = removes the RNA primer and replaces it with DNA

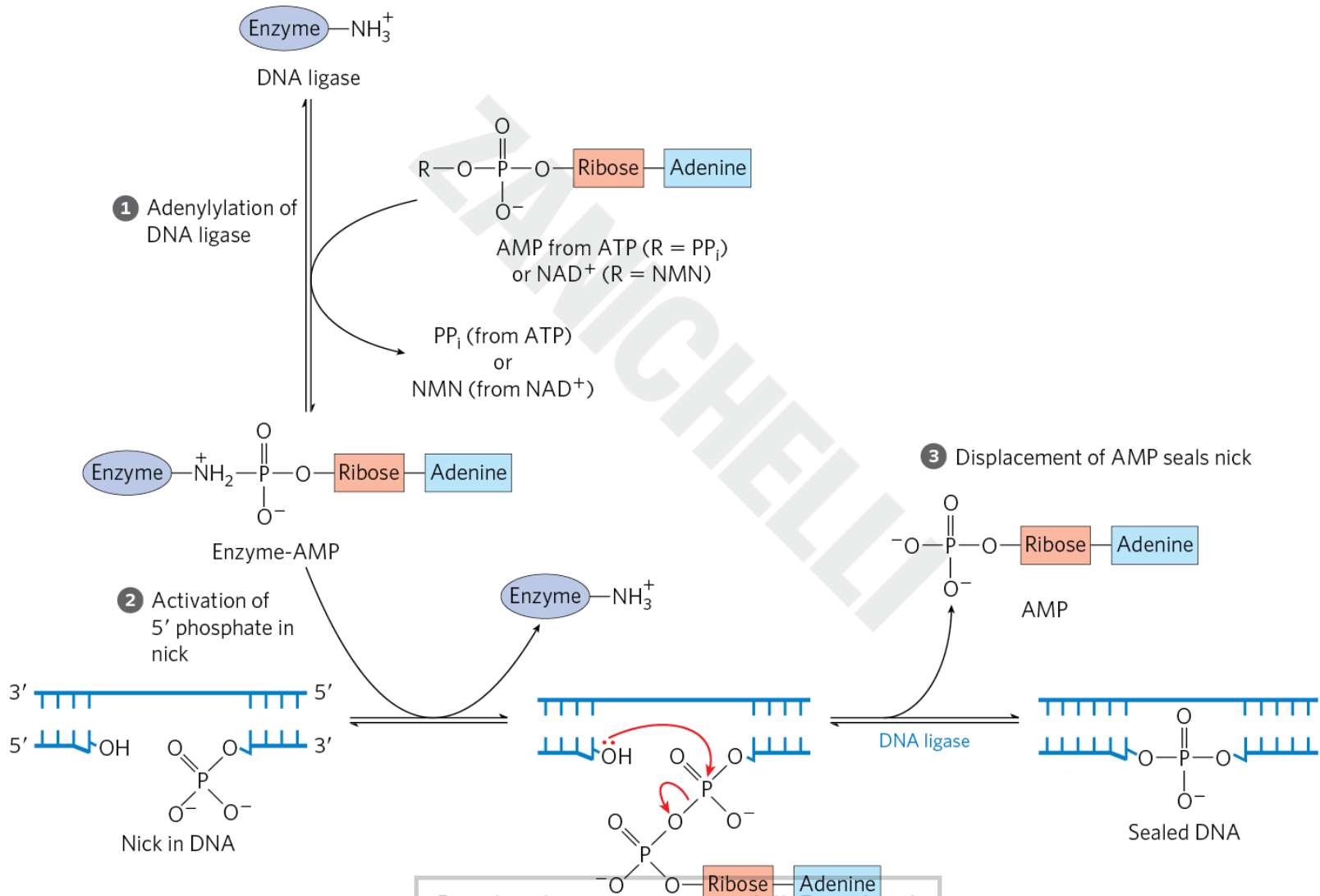


- DNA ligase = seals the remaining nick

# DNA Ligase

- DNA ligase = catalyzes the formation of a phosphodiester bond between a 3'-OH at the end of one DNA strand and a 5' phosphate at the end of another strand
  - the phosphate must be activated by adenylylation using ATP (virus and eukaryotes) or NAD<sup>+</sup> (bacteria)

# The DNA Ligase Mechanism

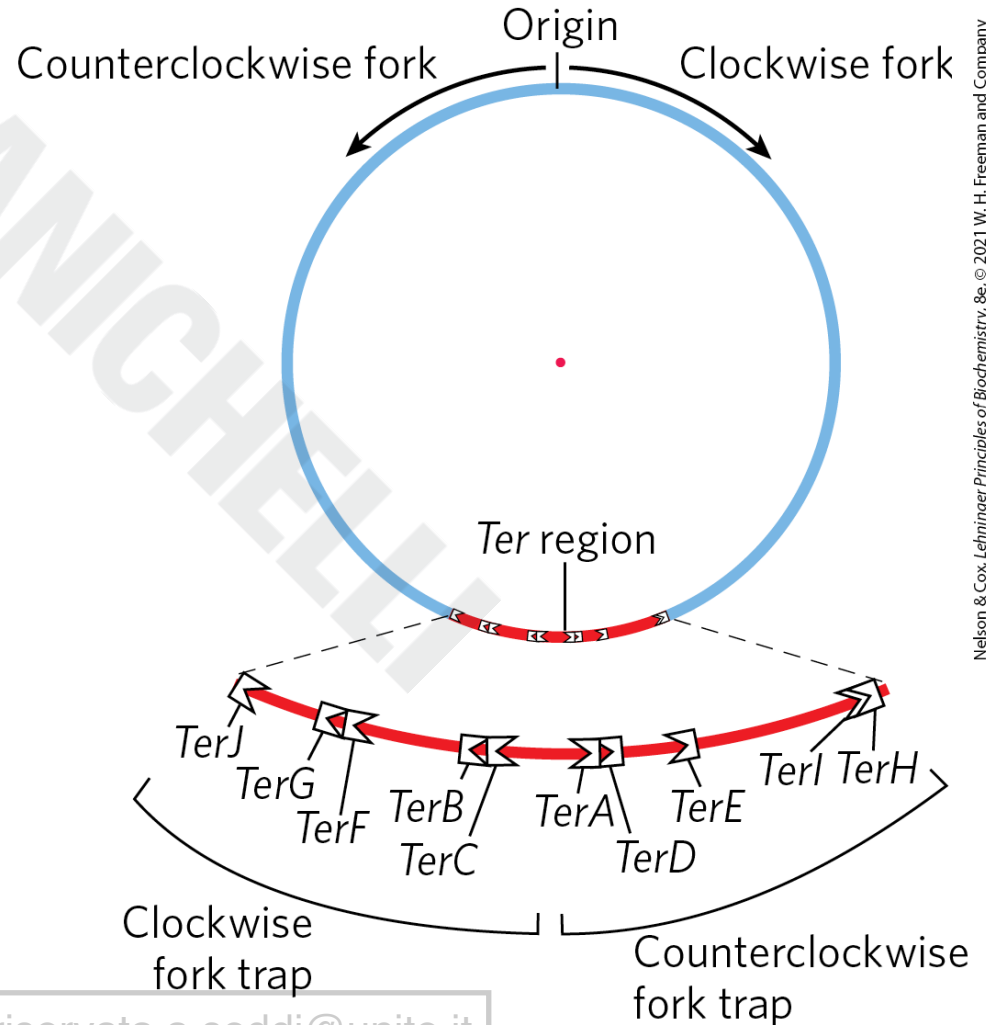


# Termination

- replication forks meet at a region with multiple copies of a 20-bp sequence called Ter
- Ter sequences are found near each other, but in opposite directions
  - creates a site that replication forks cannot leave
- Ter is also a binding site for the protein Tus (terminus utilization sequence)
  - causes a replication fork to stop

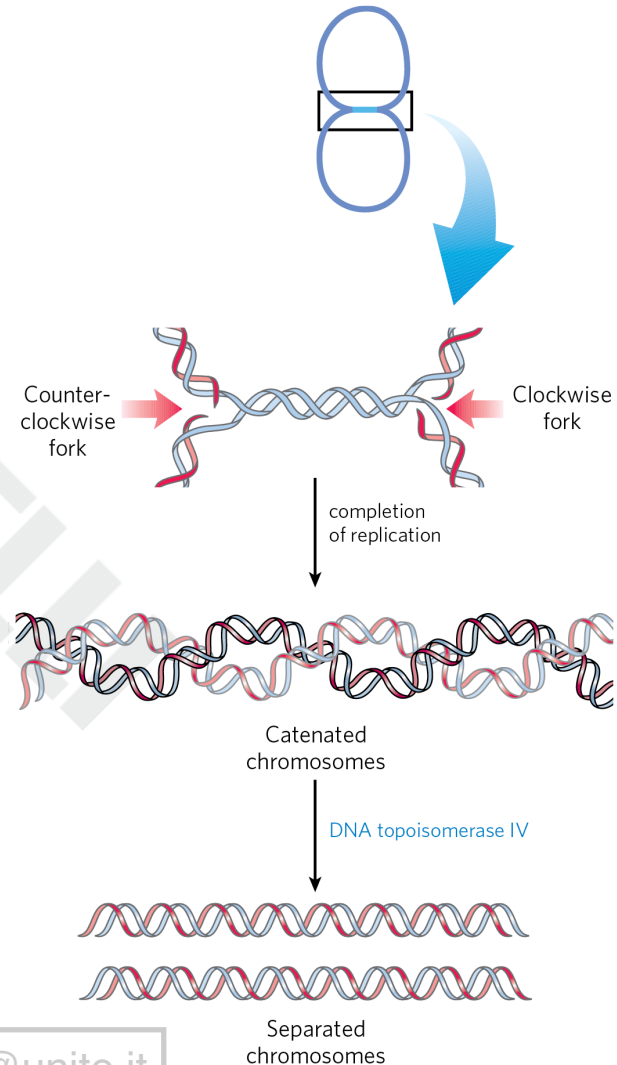
# Termination of Chromosome Replication in *E. coli*

- only one Tus-Ter complex functions per replication cycle



# Separation of Catenanes by Topoisomerase IV

- **catenanes** = two topologically interlinked circular chromosomes
- separation of the catenated circles in *E. coli* requires topoisomerase IV (a type II topoisomerase)



# Replication in Eukaryotic Cells Is Similar but More Complex

- **replicators** = yeast (*S. cerevisiae*) replication origins
  - also called autonomously replicating sequences (ARSs)
  - yeast have ~400 replicators
  - each spans ~150 bp and contains several essential, conserved sequences
- human chromosomes have ~30,000 to 50,000 replication origins

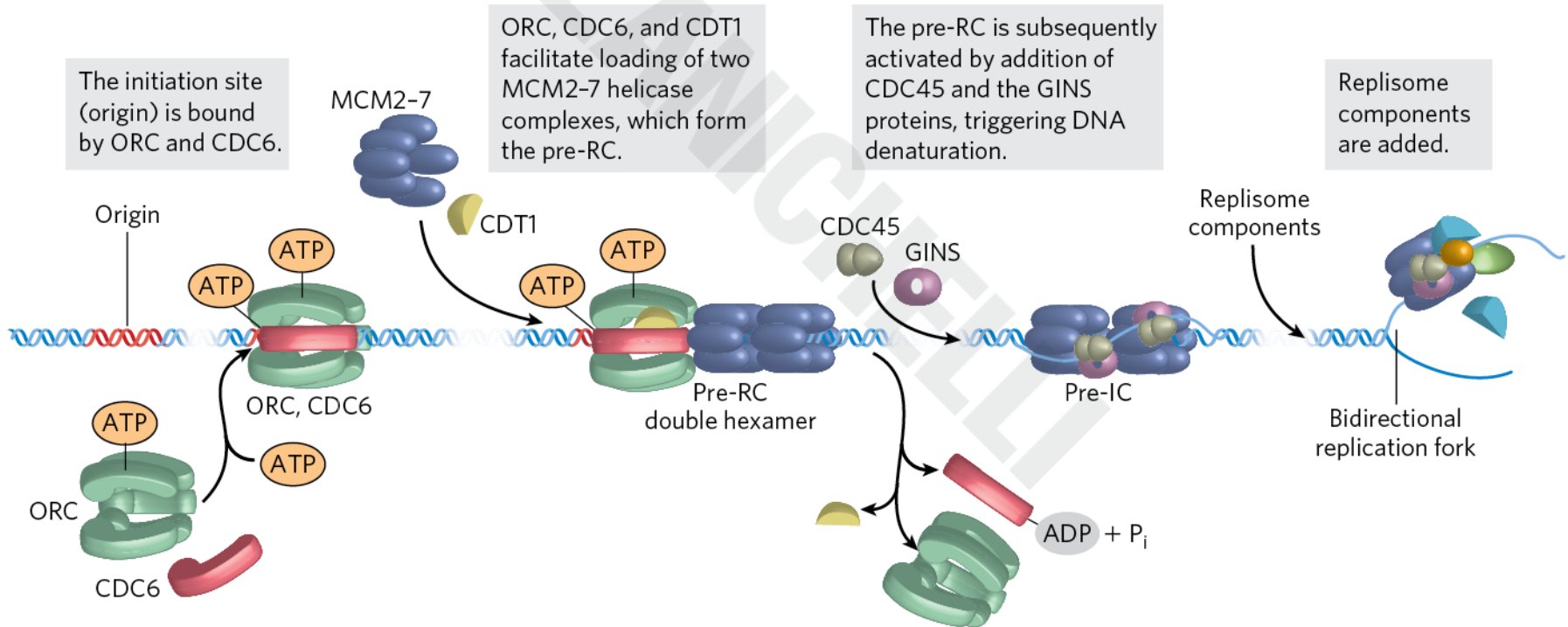
# Regulation Ensures Cellular DNA Is Replicated Once Per Cell Cycle

- regulation involves cyclins and the cyclin-dependent kinases (CDKs)
  - cyclins undergo ubiquitin-dependent proteolysis at the end of the M phase (mitosis)
- **prereplicative complexes (pre-RCs)** = established on replication initiation sites in the absence of cyclins
  - their formation (sometimes called **licensing**) renders the cell competent for replication

# Initiation of Replication in Eukaryotes

- **ORC (origin recognition complex)** = a six protein complex that recognizes and finds the origin
  - functions like DnaA
- initiation involves the loading of a replicative helicase composed of **minichromosome maintenance (MCM) proteins** (MCM2 to MCM7)
- the MCM2–7 helicase translocates 3'→5' along the leading strand template
  - similar to DnaB helicase

# Assembly of a Prereplicative Complex at a Eukaryotic Replication Origin



# Eukaryotic Rate of Replication

- the replication fork moves ~50 nucleotides/second
  - ~1/20th the rate seen in *E. coli*
- eukaryotes compensate for the slower rate by having many origins

# Eukaryotes Have Several Types of DNA Polymerases

- **DNA polymerase  $\epsilon$**  = synthesizes the leading strand
  - highly processive
  - has 3'→5' proofreading exonuclease activity
- **DNA polymerase  $\delta$**  = synthesizes the lagging strand
  - has 3'→5' proofreading exonuclease activity
- **DNA polymerase  $\alpha$**  = a DNA polymerase/primase that synthesizes RNA primers and also extends them to initiate synthesis of each Okazaki fragment
  - does not have 3'→5' proofreading exonuclease activity

# Function of PCNA

- proliferating cell nuclear antigen (PCNA) = a protein that forms a circular clamp to enhance the processivity of two polymerases
  - analogous function to the  $\beta$  subunit

# Functions of RPA and RFC

- RPA (replication protein A) = single-stranded DNA-binding protein
  - equivalent in function to the *E. coli* SSB protein
- RFC (replication factor C) = a clamp loader for PCNA that facilitates the assembly of active replication complexes
  - significant sequence similarity to the subunits of the bacterial clamp-loading ( $\gamma$ ) complex

# Termination of Eukaryotic Replication

- termination occurs when replication forks operating from nearby origins converge

# Viral DNA Polymerases Provide Targets for Antiviral Therapy

- acyclovir = inhibits the DNA polymerase of the herpes simplex virus
  - converted to acyclo-GTP
- acyclo-GTP competitively inhibits viral DNA polymerases and terminates replication

